

# Comparative Evaluation of Ozone Treatment in Critical Size Bone Defects Reconstructed with Alloplastic Bone Grafts

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**How to cite this paper:** Laçın, N., Kaya, B., Deveci, E., Kadiroğlu, E.T., Aktaş, A., Yalçın, M. and Uysal, E. (2018) Comparative Evaluation of Ozone Treatment in Critical Size Bone Defects Reconstructed with Alloplastic Bone Grafts. *International Journal of Clinical Medicine*, 9, 566-579.

<https://doi.org/10.4236/ijcm.2018.97048>

**Received:** June 5, 2018

**Accepted:** July 17, 2018

**Published:** July 20, 2018

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## Abstract

The purpose of this study was to investigate osteogenesis promoted by osteoconductive properties of bone grafting materials and the histopathological effects of ozone on osteogenesis. In total, 56 Wistar rats were equally divided into 4 groups. In control group, calvarial bone defect was created in 14 rats. For second group, 8 mm calvarial bone defect with ozone treatment was applied in 14 rats. For third group, an alloplastic bone graft was implanted on 8 mm calvarial bone defect. In fourth group, alloplastic bone graft was inserted in calvarial defect and ozone was treated additionally. Seven of the rats were sacrificed at the end of 4th week and the remaining 7 were sacrificed at the end of 8th week of experiment. In the study, the periosteal flaps were removed with a thin periosteal elevator and averagely 0.8 cm diameter-circular full bone defect was created with a specially designed trephine drill. The bone from the calvarial region was fixed in 10% formalin solution. After decalcification, bones were taken for routine paraffin blocking. Sections were stained with Hematoxylin-Eosin and Masson Trichrome. Histopathological findings of 4th and 8th weeks rats showed that best result for new bone formation was observed in graft + ozone treatment. It is concluded that ozone treatment increases the hemostasis in graft region, induces angiogenesis, promotes cell proliferation by preventing infiltration, induces matrix formation by influencing osteoblastic activity and has a positive effect in osteogenesis.

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## Keywords

Calvarial Bone Graft, Ozone, Osteoblast Cells, Rat

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### 1. Introduction

Fracture healing depends on many actors: Local factors such as the degree of trauma, the position of the wound edges relative to each other, the vascularization of the wound area, the type of bone, the degree of immobilization, the amount of local necrosis and soft tissue damage, the presence of infections and systemic diseases (*i.e.* age, diabetes, anemia, tuberculosis); hormones such as parathyroid, calcitonin, insulin, growth hormones; vitamins such as A, C and D and general factors such as chondroitin sulphate and exercises [1] [2]. Cortical grafts provide a durable and rigid structure but they have no ability to increase osteogenesis. The primary advantage of cancellous bone and bone marrow is that they are able to significantly enhance osteogenesis. These abilities depend on the fact that they have viable cells that can transform into osteoblasts as well as those that induce osteogenesis. The only known disadvantage of these grafts is that they can't provide mechanical stability [3] [4].

Allografts are bone tissues obtained from genetically different individuals but sharing properties of same species with donor. Fresh frozen bone can be classified as frozen dried bone and demineralized bone matrix [5] [6]. Due to the limited availability of autografts, undesirable features of allografts and xenografts such as the risk of disease transfer, researchers have now been focused on synthetically graft materials which of them have been produced for use in bone defects. Since large numbers of those materials are produced, they should be studied well in researches. Alloplasts have become a necessary material in recent years for the repair of maxillofacial skeleton [7] [8].

Ozone is available in the stratosphere layer of earth at a concentration of 1 to 10 ppm in the gas phase. The disinfection property of ozone is due to its strong oxidizing property. Not only do they kill viruses and bacteria, they can also oxidize all microorganisms and their toxins. Ozone can also effectively neutralize phenols, pesticides, detergents, chemical wastes, and aromatic compounds [9]. Ozone is widely used in stomatology as a disinfectant. Besides, it also can be used in wound healing after tooth extraction, the treatment of oral infections, cases of candidiasis or periodontitis, stomatitis, treatment of infected wounds in soft tissues, and oral cavity washing prior to surgery [10]. Ozone treatments have positive effects in studies of on wound healing, ischemic and infectious diseases. It is also effectively used in many infectious diseases from simple dental and oral infections to hepatitis [11]. Studies show that ozone treatment has benefited significantly in pit and fissure caries [12], non-cavitated occlusal caries [13], and root caries at the initial stage [12] [13] [14]. The aim of this study was to investigate the histopathological effects of ozone on bone graft materials and to ex-

amine osteogenesis development induced by osteoconductive properties of these materials.

## 2. Materials and Methods

This study was carried out at the Dicle University Health Sciences and Application Center (DUSAM) with the approval of the Dicle University Experimental Animal Ethics Committee with the protocol dated 12.11.2013 and numbered 2013/55. In the study, 56 male Wistar rats weighing 280 - 300 grams were used and housed individually in suitable cages at temperatures of  $22^{\circ}\text{C} \pm 2^{\circ}\text{C}$  and 12 hours of dark/12 hours of light conditions. The nutritional needs of the animals were met regularly with standard laboratory food and water. Four groups (14 rats per group) were arranged as below:

**1) Control Group:** Calvarial bone defect was created in 14 rats without any additional treatment and wound was sutured. Seven of the subjects were sacrificed at the end of the 4th week, and the remaining 7 were sacrificed at the end of the 8th week.

**2) Experimental Group:** 8 mm calvarial bone defects were created in all rats and then treated with ozone. 7 of the subjects were sacrificed at the end of the 4th week, and the remaining 7 were sacrificed at the end of the 8th week.

**3) Experimental Group:** 8 mm calvarial bone defects were created in all rats and then alloplastic bone grafts were applied to the defect. 7 of rats were sacrificed at the end of the 4th week and the remaining were sacrificed at the end of the 8th week.

**4) Experimental Group:** 8 mm calvarial bone defects were created in all rats and then alloplastic bone grafts + ozone treatment was applied to the defect. Seven of them were sacrificed at the end of the 4th week, and the remaining were sacrificed at the end of the 8th week.

### 2.1. Surgical Operation

The animals were anesthetized with intraperitoneally 3 mg/kg xylazine (Rompun 2%; Bayer) and 90 mg/kg Ketamine HCl (Ketalar; Eczacıbaşı-Warner Lambert). Skin was incised to open frontal bone. A periosteal flap was removed with a thin elevator. Surgical sites were exposed with an incision through the skin and the periosteum at the midline of the calvaria. The periosteal flap was removed with a thin periosteal elevator and a specially designed trephine bur was created with a circular full-thickness bone defect with a diameter of 0.8 cm on the midline. After the graft was applied in our study, topical Ozone (Prozone W & H, Bürmoos, Austria) was applied for 12 seconds with a Coro-type applicator. Allografts are bone tissues of the same species as the recipient but derived from genetically distinct individuals. It is used as fresh frozen bone, frozen dried bone and demineralized bone matrix. Allograft material was used as demineralized bone matrix in our study.

Alloplastic material (Bio-Graft-HT) was placed in defect area in group 3 and group 4. Subcutaneous tissue was sealed with 6/0 vicryl suture and skin was

closed using 5/0 silk suture. Half of the experimental animals in all groups were sacrificed at the end of the 4th week and the remainder was sacrificed at the end of the 8th week by intraperitoneally overdose of sodium thiopentone. The skin on the calvarium was completely removed and the defect was taken out with bone forceps.

## 2.2. Histopathological Protocol

The bones from the calvarial region were fixed in 10% formalin solution. They were decalcified for 1 week in 10% ethylene diamine tetra acetic acid (EDTA) solution. After washing, tissues were dehydrated through ascending alcohol series and cleared with xylene. After embedding them in paraffin, 4 - 6  $\mu\text{m}$  thick sections were taken with Leica ultramicrotome. Sections were stained with Masson Trichrome and Hematoxylin-Eosin and examined by light microscope.

## 2.3. Statistical Analysis

Statistics and analyzes were performed using the SPSS 22.0 for Windows computer package program, and the results were considered statistically significant for  $p < 0.05$ . In the analysis of the data, Kruskal-Wallis and Mann-Whitney U non-parametric statistical tests were used in the intergroup comparisons depending on the variables and the results were given as mean and standard deviation values.

## 3. Results

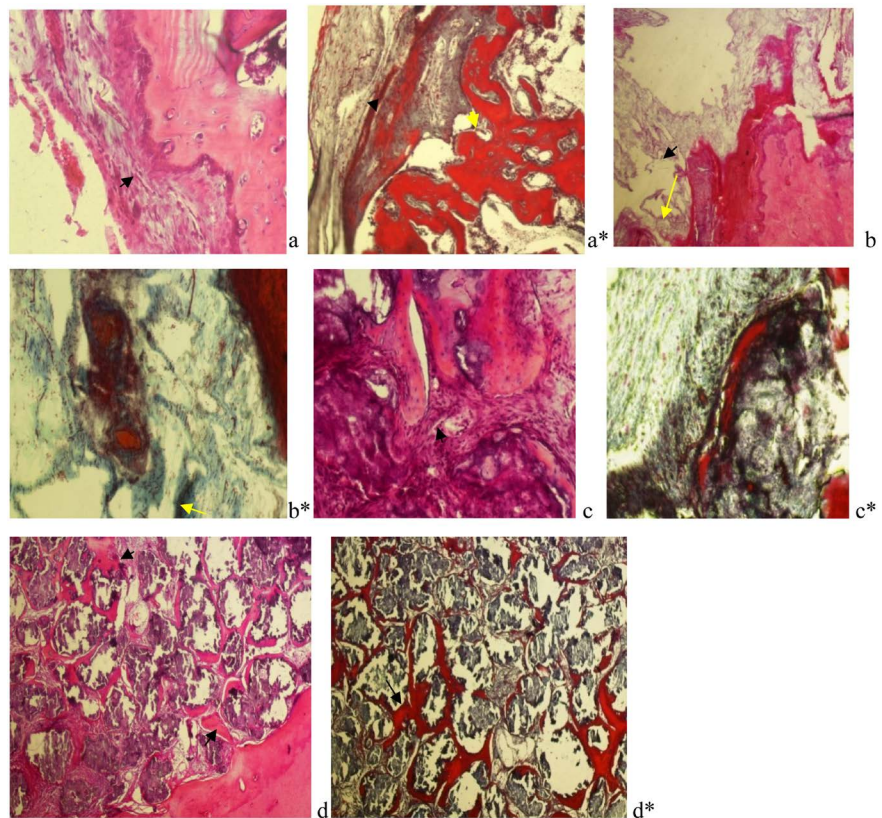
### Histopathological Findings

#### 1) 4th week findings

**a) Control Group:** **Figure 1(a)** shows a significant increase was observed in osteoclast cells in defect area. Hemorrhage and dilatation in blood vessels and mononuclear cell infiltration were increased. Defect Group: In **Figure 1(a\*)**, in other section of the same group, an increase in fibrous tissue and collagen fibers, freely dispersed erythrocytes due to tearing in the vessel wall, intense mononuclear cell infiltration and rarely edema were observed.

**b) Defect + Ozone treated Group:** Sections showed dilatation was significantly increased in the veins. Comparing with the defect group, there was a decrease in fibrous tissue and osteoblastic activity in this group. In another section, the collagen fibers were bundled, the connective tissue were increased, new bone trabeculae were formed and the bone matrix began to become evident.

**c) Defect + Graft Applied Group:** Osteoblasts were clearly observable, connective tissue cells and fibrous structure around bone trabeculae were increased, graft resorption was decreased. A decrease in osteoblasts and cell infiltration was observed. In another section, bone trabeculae were more abundant around the veins and collagen fibers were observed in bundles. Decrease in connective tissue cells and dispersed erythrocytes were observed. A small number of osteocytes were found in bone trabeculae.



**Figure 1.** a: A significant increase was observed in osteoblast cells in defect area (arrow). H-E staining Bar 50  $\mu$ m; a\*: An increase in fibrous tissue and collagen fibers (arrow), dilatation in blood vessels (yellow arrow) Trichrome Masson staining Bar 50  $\mu$ m, b-Inflamatory cells in defect area (arrow) H-E staining Bar 50  $\mu$ m; b\*: hemorrhage in blood vessels (yellow arrow) Trichrome Masson staining Bar 50  $\mu$ m, c-osteoblast cells around bone trabecula (arrow), H-E staining Bar 50  $\mu$ m; c\*: intense inflammatory cells in graft area (yellow arrow) Trichrome Masson staining Bar 50  $\mu$ m; d: An increase in osteoblastic activity and osteocyte cells (arrow), H-E staining Bar 50  $\mu$ m; d\*: Appearance of mature bone particles (arrow) Trichrome Masson staining Bar 50  $\mu$ m.

**d) Defect + Graft + Ozone Applied Group:** In sections of this group, graft resorption began, bone trabeculae became evident due to an increase in dense connective tissue fibers and osteoblastic activity around the graft. Matrix development was increased and a small number of osteocytes in bone trabeculae were observed. In another section, dense connective tissue around the graft was enriched by especially collagen fibers, new bone formation in connective tissue began to increase and osteocyte cells with their lacunae became evident in new bone trabeculae.

For the 4<sup>th</sup> week, cell infiltration was significantly different between the groups ( $p < 0.0001$ ). When we examined osteoclastic activity of week 4, there was a significant difference between the groups ( $p = 0.0001$ ). There was a significant difference between the groups in osteoblastic activity of 4th week ( $p = 0.253$ ). There was a significant difference between the groups in terms of matrix formation ( $p = 0.0007$ ), bone trabeculae ( $p = 0.0018$ , vascular dilatation and hemorrhage ( $p =$

0.0005) and collagen fiber distribution ( $p = 0.0099$ ) (Table 1).

## 2) 8th week findings

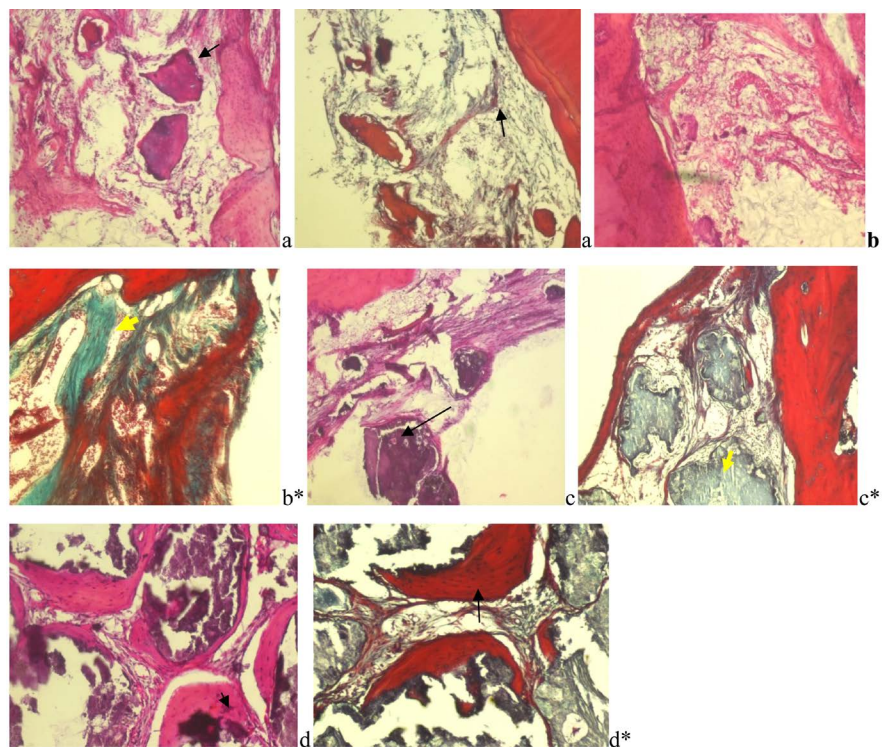
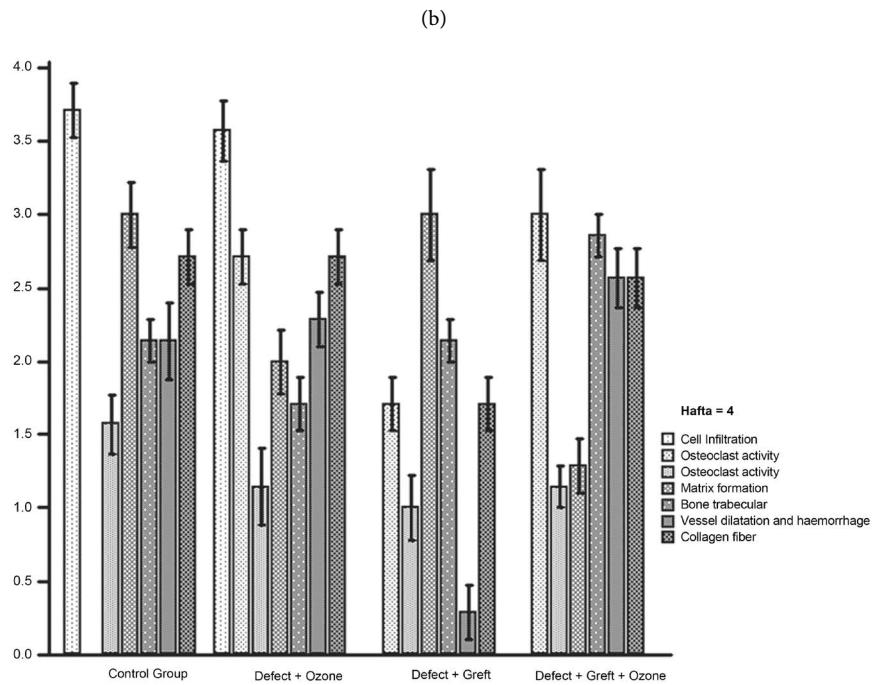
**a) Control Group:** In Figure 2(a), bone trabeculae were visible and osteoblasts were flattened and arranged in a single row around the trabeculae. While cell infiltration mostly gathered around the trabeculae, edema was seen in some areas. Vessels were dilated and apparently hemorrhagic. Figure 2(a\*) shows the collagen fibers were in bundled form, islets of bone trabeculae were seen, the osteoblastic activity was increased in the trabeculae and the matrix development

**Table 1.** Statistical evaluation of parameters according to 4th week according to different groups.

(a)						
Parameter's	Groups	n	Mean	Average Rank	Kruskal-Wallis Test value	Multiple comparisons for groups ( $p < 0.05$ )
Cell infiltration	(1) Control group	7	3.71	22.00	20.3276; $p < 0.0001$	(3) (4)
	(2) Defect + Ozone	7	3.57	21.00		(3) (4)
	(3) Defect + Graft	7	0.00	7.50		(1) (2)
	(4) Defect + Graft + Ozone	7	0.00	7.50		(1) (2)
Osteoclastic activity	(1) Control group	7	0.00	4.00	20.1291; $p = 0.0001$	(2) (3) (4)
	(2) Defect + Ozone	7	2.71	20.07		(1) (3)
	(3) Defect + Graft	7	1.71	12.43		(1) (2) (4)
	(4) Defect + Graft + Ozone	7	3.00	21.50		(1) (3)
Osteoblastic activity	(1) Control group	7	1.57	18.93	2.9008; $p = 0.2533$	ns
	(2) Defect + Ozone	7	1.14	13.79		
	(3) Defect + Graft	7	1.00	11.93		
	(4) Defect + Graft + Ozone	7	1.14	13.36		
Matrix formation	(1) Control group	7	3.00	20.50	15.5120; $p = 0.0007$	(2) (4)
	(2) Defect + Ozone	7	2.00	11.71		(1) (3) (4)
	(3) Defect + Graft	7	3.00	20.00		(2) (4)
	(4) Defect + Graft + Ozone	7	1.29	5.79		(1) (2) (3)
Bone trabeculae	(1) Control group	7	2.14	13.36	10.6784; $p = 0.0018$	(4)
	(2) Defect + Ozone	7	1.71	8.64		(4)
	(3) Defect + Graft	7	2.14	13.36		(4)
	(4) Defect + Graft + Ozone	7	2.86	22.64		(1) (2) (3)
Vessel dilatation and hemorrhage	(1) Control group	7	2.14	16.29	15.6492; $p = 0.0005$	(3)
	(2) Defect + Ozone	7	2.29	17.36		(3)
	(3) Defect + Graft	7	0.29	4.14		(1) (2) (4)
	(4) Defect + Graft + Ozone	7	2.57	20.21		(3)
Collagen fibers	(1) Control group	7	2.71	17.79	9.0654; $p = 0.0099$	(3)
	(2) Defect + Ozone	7	2.71	17.79		(3)
	(3) Defect + Graft	7	1.71	6.50		(1) (2) (4)
	(4) Defect + Graft + Ozone	7	2.57	15.93		(3)

ns: non-significant;  $p > 0.05$ .





**Figure 2.** a: bone trabeculae in defect area (arrow). H-E staining Bar 50  $\mu$ m; a\*: An increase in fibrous tissue and collagen fibers(arrow), Trichrome Masson staining Bar 50  $\mu$ m; b: Inflammatory cells in defect area (arrow) H-E staining Bar 50  $\mu$ m; b\*: increase of collagen fibers in defect area (yellow arrow) Trichrome Masson staining Bar 50  $\mu$ m; c: Appearance of mature bone trabecula (arrow), H-E staining Bar 50  $\mu$ m; c\*: Enlargement bone trabecula in graft area (yellow arrow) Trichrome Masson staining Bar 50  $\mu$ m; d: An increase in bone trabeculae and osteocyte cells (arrow), H-E staining Bar 50  $\mu$ m; d\*: Appearance of mature bone particles (arrow) Trichrome Masson staining Bar 50  $\mu$ m.

started. Extracellular matrix of connective tissue was increased. Form of fully developed mature bone cells were not observed in this group.

**b) Defect + Ozone Applied Group:** Islets of inflammatory cells; hemorrhage and dilatation in vessels of the defect area can be seen in **Figure 2(b)**. Throughout new bone formation, matrix development was apparently begun. There were bone trabeculae in some areas. **Figure 2(b\*)** shows thickening in collagen fibers, intense hemorrhagic areas in defect region, enlargement of the bone matrix and new bone trabeculae formation were observed.

**c) Defect + Graft Group:** The histological examination of this group showed new bone trabeculae with increased connective tissue around the graft sites. As cell infiltration continued, freely distributed erythrocytes were found in connective tissue. Osteoblast cells were clearly observed around the bone trabeculae as seen in **Figure 2(c)**. Graft resorption was dispersed and graft areas began to diminish gradually. In **Figure 2(c\*)**, collagen fibers were tightly gathered around graft areas and made resorbed grafts reduce. Inflammatory cells spread diffusely between graft areas, and new bone particles began to become apparent in the periphery of graft.

**d) Defect + Graft + Ozone Applied Group:** **Figure 2(d)** shows that the new bone trabeculae around the graft areas began to turn into mature flat bone and osteocytes cells were abundant due to increased osteoblastic activity. Cell infiltration was gradually reduced and mature bone trabeculae were formed in the site of resorbed graft. In **Figure 2(d\*)**, revealed shrinkage in the resorbed graft sites, the structures of mature bone particles became evident and fibrillary development in the connective tissue accelerated and wrapped around the bone trabeculae.

There was a significant difference between the groups when the weekly cell infiltration ( $p = 0.0014$ ), osteoclastic ( $p = 0.0004$ ) and osteoblastic ( $p = 0.0055$ ) activities were examined. Matrix formation at 8 weeks ( $p = 0.0001$ ), Bone trabeculae ( $p = 0.0002$ ) There was a significant difference between the groups when vascular dilatation and hemorrhage ( $p < 0.0001$ ) and collagen fiber distribution ( $p < 0.0001$ ) (**Table 2** and **Table 3**).

**Table 2.** Statistical evaluation of parameters according to 8th week according to different groups.

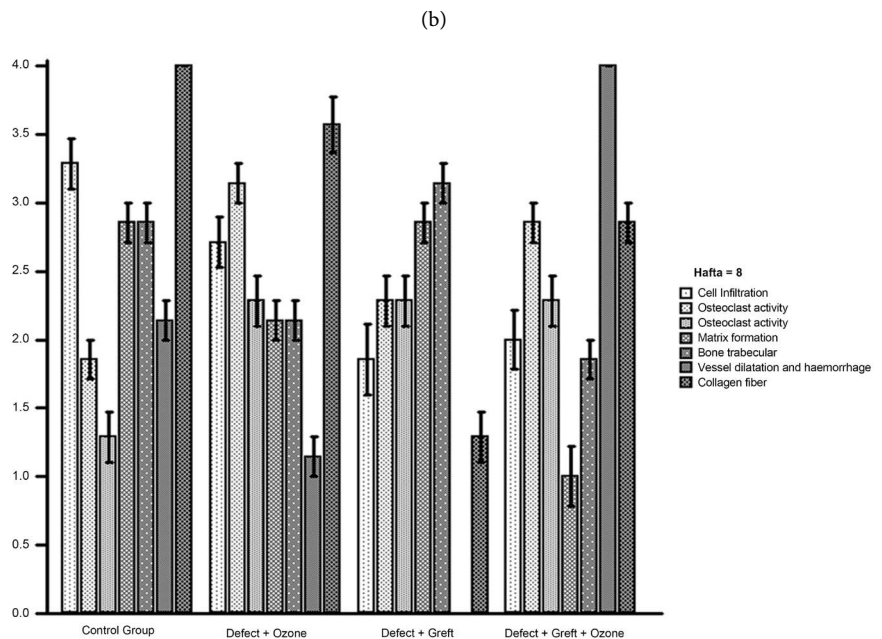
(a)						
Parameter	Groups	n	Mean	Average Rank	Kruskal-Wallis Test value	Multiple comparisons for groups ( $p < 0.05$ )
Cell infiltration	(1) Control	7	3.29	22.50	13.3723; $p = 0.0014$	(3) (4)
	(2) Defect + Ozone	7	2.86	17.21		(3) (4)
	(3) Defect + Graft	7	2.86	8.64		(1) (2)
	(4) Defect + Graft + Ozone	7	1.14	9.64		(1) (2)
Osteoclastic activity	(1) Control	7	2.71	6.57	14.5683; $p = 0.0004$	(2) (4)
	(2) Defect + Ozone	7	1.29	21.57		(1) (3)



Continued

	(3) Defect + Graft	7	1.00	11.21		(2) (4)
	(4) Defect + Graft + Ozone	7	0.00	18.64		(1) (3)
	(1) Control	7	1.86	6.14		(2) (3) (4)
Osteoblastic activity	(2) Defect + Ozone	7	2.29	17.29	9.6334; $p = 0.0055$	(1)
	(3) Defect + Graft	7	2.86	17.29		(1)
	(4) Defect + Graft + Ozone	7	4.00	17.29		(1)
	(1) Control	7	2.00	20.43		(2) (4)
Matrix formation	(2) Defect + Ozone	7	2.29	12.57	17.8543; $p = 0.0001$	(1) (3) (4)
	(3) Defect + Graft	7	2.14	20.43		(2) (4)
	(4) Defect + Graft + Ozone	7	4.00	4.57		(1) (2) (3)
	(1) Control	7	1.86	19.14		(2) (4)
Bone trabeculae	(2) Defect + Ozone	7	2.29	9.86	16.0978; $p = 0.0002$	(1) (3)
	(3) Defect + Graft	7	3.14	22.00		(2) (4)
	(4) Defect + Graft + Ozone	7	3.57	7.00		(1) (3)
	(1) Control	7	3.14	17.57		(2) (3) (4)
Vessel dilatation and hemorrhage	(2) Defect + Ozone	7	2.86	11.43	24.7621; $p < 0.0001$	(1) (3) (4)
	(3) Defect + Graft	7	1.86	4.00		(1) (2) (4)
	(4) Defect + Graft + Ozone	7	1.29	25.00		(1) (2) (3)
	(1) Control	7	2.29	23.00		(2) (3) (4)
Collagen fibers	(2) Defect + Ozone	7	2.14	18.71	20.9831; $p < 0.0001$	(1) (3) (4)
	(3) Defect + Graft	7	2.14	4.14		(1) (2) (4)
	(4) Defect + Graft + Ozone	7	2.86	12.14		(1) (2) (3)

ns: non-significant;  $p > 0.05$ .



**Table 3.** Statistical evaluation of parameters according to 4th and 8th week according to different groups.

Parameters	Groups	Weeks	Mean	Mean Rank	<i>p</i>	
Cell infiltration	Control group	4	3.71	1.71	0.180	
		8	3.29	1.29		
	Defect + Ozone	4	3.00	1.57	0.655	
		8	2.86	1.43		
	Defect + Graft	4	3.00	1.57	0.655	
		8	2.86	1.43		
	Defect + Graft + Ozone	4	2.29	2.00	0.008	
		8	1.14	1.00		
	Osteoclastic activity	Control group	4	3.57	1.93	0.014
			8	2.71	1.07	
Defect + Ozone		4	1.57	1.64	0.317	
		8	1.29	1.36		
Defect + Graft		4	1.29	1.57	0.317	
		8	1.00	1.43		
Defect + Graft + Ozone		4	0.29	1.64	0.157	
		8	0.00	1.36		
Osteoblastic activity		Control group	4	0.00	1.00	0.008
			8	1.86	2.00	
	Defect + Ozone	4	1.14	1.07	0.014	
		8	2.29	1.93		
	Defect + Graft	4	2.14	1.14	0.025	
		8	2.86	1.86		
	Defect + Graft + Ozone	4	2.57	1.00	0.008	
		8	4.00	2.00		
	matrix formation	Control group	4	.00	1.00	0.008
			8	2.00	2.00	
Defect + Ozone		4	1.00	1.00	0.008	
		8	2.29	2.00		
Defect + Graft		4	1.71	1.29	0.083	
		8	2.14	1.71		
Defect + Graft + Ozone		4	2.71	1.00	0.008	
		8	4.00	2.00		
Bone trabeculae		Control group	4	0.00	1.00	0.008
			8	1.86	2.00	
	Defect + Ozone	4	1.14	1.07	0.014	
		8	2.29	1.93		
	Defect + Graft	4	2.14	1.07	0.014	
		8	3.14	1.93		
	Defect + Graft + Ozone	4	2.71	1.14	0.025	
		8	3.57	1.86		

## Continued

Vessel dilatation and hemorrhage	Control group	4	2.71	1.29	0.083
		8	3.14	1.71	
	Defect + Ozone	4	3.00	1.57	0.564
		8	2.86	1.43	
	Defect + Graft	4	2.86	1.93	0.014
		8	1.86	1.07	
Defect + Graft + Ozone	4	1.71	1.71	0.083	
	8	1.29	1.29		
Collagen fibers	Control group	4	1.71	1.21	0.046
		8	2.29	1.79	
	Defect + Ozone	4	2.00	1.43	0.564
		8	2.14	1.57	
	Defect + Graft	4	2.14	1.50	1.000
		8	2.14	1.50	
Defect + Graft + Ozone	4	2.57	1.36	0.317	
	8	2.86	1.64		

When compared histologically at 4 and 8 weeks; colloid fiber growth, matrix formation and new bone formation were less than 4 weeks to 8 weeks. Vessel dilatation and hemorrhage increased cell infiltration over 4 weeks to 8 weeks, osteoblastic activity significantly increased bone trabecular formation at 8 weeks. In bone formation, post-defect graft application was evident at 8 weeks, ozone application increased bone formation as an inducing agent.

#### 4. Discussion

Nagayoshi *et al.* [15] found that no microorganisms remained in the plaque after ozonized water treatment on the dental plaque. In oral surgery, ozonized water is used to provide hemostasis, to increase local oxygen delivery to the site, and to inhibit bacterial proliferation [13]. Ozonized water has also been proposed to use as a prophylactic agent against infection after osteotomy during oral surgery. Following the high dose of radiotherapy treatment, ozone treatment has been shown to be effective in improvement of wound healing [16]. After radiotherapy in maxilla or mandible, in areas where especially oxygen is greatly reduced, spongy medullary regions become inadequately vascularized due to deterioration of vascularity, resulting in aseptic osteonecrosis [16]. In surgical procedures such as tooth extraction or implant application, the healing of the damaged region takes longer than the healing of the healthy bone [16] [17]. Schmitz and his colleagues studied in 3, 4, 5 and 8 mm defects and suggest that an 8 mm defect is an ideal defect [18]. In our study, 8 mm defects were created. Different bone graft materials have been used for bone regeneration, closure of osteotomy openings, and alveolar augmentation in oral and maxillofacial surgeons [19]

[20]. Natural coral-derived grafts, synthetic bone graft materials, are used in alveolar crest elevation, intra-bone defects, material loss fractures, facial bone defects, orthognathic surgery, and maxillary sinus ground [20] [21] [22]. In our study, an alloplastic graft material consisting of a combination of 350 - 500  $\mu\text{m}$ -diameter porous biphasic hydroxyapatite granules and  $\beta$ -tricalcium phosphate granules was used. Agrillo *et al.* have studied the effects of ozone on wound healing after tooth extraction in 15 patients using bisphosphonates. Topical ozone gas with  $\beta$ -lactam antibiotics were applied to all patients 1 week before and after tooth extraction. Clinical and radiological follow-ups of patients showed complete healing in the extraction socket [23]. Kazancıoğlu *et al.* formed a 5 mm diameter bone defect in 30 Wistar rats. They created defects and filled them with biphasic calcium phosphate grafts. They categorized rats into three groups and histomorphometrically compared the effect of laser and ozone on new bone formation. Kazancıoğlu *et al.* observed significant bone formation in both groups compared to the control group [24]. Özdemir *et al.* have studied the effect of ozone treatment on autogenous bone graft healing in calvarial defects. Histological results of this study revealed that there was an increase in osteoblastic activity and significant difference in bone formation in ozone treated group [25]. In the histopathological examination of week 4 of our study, a significant increase in osteoblastic activity was observed in all compared to the control group. Week 8 results showed significant increase in osteoblastic activity in graft + ozone group compared to all other groups. More bone trabecular formation was observed in the groups treated with grafts and grafts + ozone compared to the group treated with only ozone. 4th week histological evaluations on matrix formation showed a significant increase in all groups compared with the control group at the end of. Statistically speaking on histological results, 4-week ozone treatment is less effective than 8-week ozone treatment and additionally 8-week treatment is more effective and significant than 4-week-treatment in total. In our study, there was a significant difference between the groups treated with s graft + ozone and the other groups in terms of cell infiltration in the groups sacrificed at 8 weeks. There was no significant difference in comparison between the other groups. Erdemci *et al.* studied on the effect of systemic ozone on alveolar bone healing. A total of 112 Wistar rats were used, and they were sacrificed on days 14 and 28. They reported systemic ozone treatment significantly improved alveolar bone healing after extraction [26]. In their study, unlike the half of the rats at the end of 4 week, the remaining half was sacrificed at the end of the 8th week. Each group was divided into 2 subgroups. A critical size bone defect of 8 mm in diameter was formed in the calvarium of all rats. Topical ozone was applied to the ozone treated groups for 12 sec.

In our study, it was determined that ozone application accelerated bone healing in parallel with this study. It was observed that topical ozone administration at week 4 failed to exert only osteoblastic activity on graft application. However, ozone and graft combination may be beneficial. When the bone trabeculae were

taken to evaluate the 8th week groups, a statistically significant increase was observed only in the graft and graft + ozone treated groups compared to the control group. When the 4- and 8-week findings were evaluated; the best result for new bone formation was obtained from the graft + ozone treated group. It is thought that ozone application increases the hemostasis in the region and induces angiogenesis, increases cell proliferation by preventing infiltration. It also promotes osteoblastic activity and matrix formation and has a positive effect on estrogen.

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